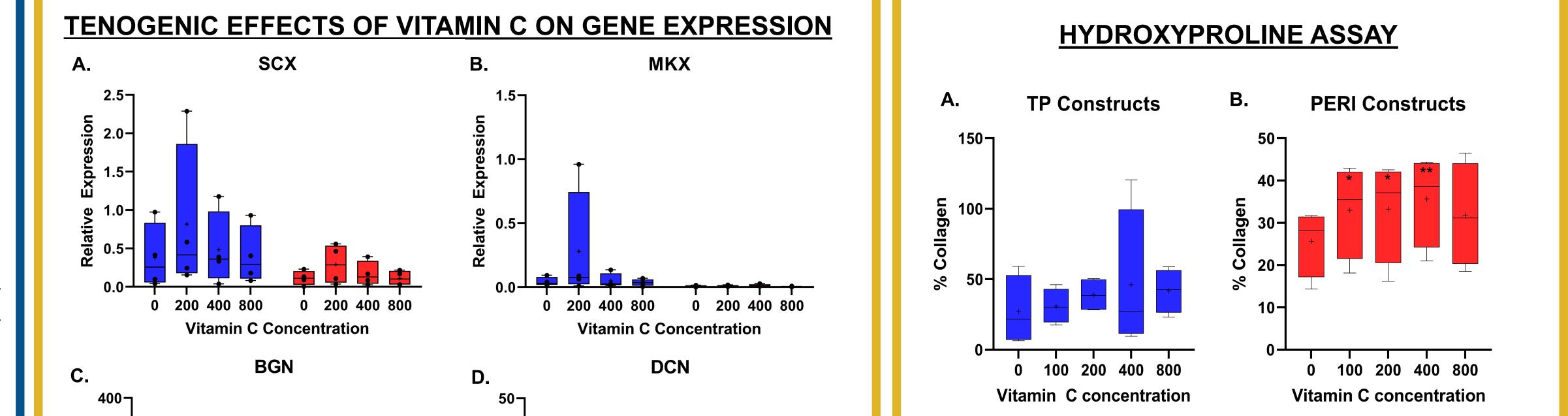


Understanding the Tenogenic and Cytoprotective Roles of Vitamin C

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BACKGROUND:

Tendons function to transfer mechanical loads from muscle to bone [1]. Tendinopathy is an overarching term that describes a wide variety of tendon diseases and their respective severities [2]. Tendinopathy of the superficial digital flexor tendon can result in debilitating lameness for horses regardless of riding discipline [3,4]. Research continues to elucidate the complex mechanism of healing that occurs after disease or injury, however there are still no curative therapies for tendinopathies. Additionally, drugs utilized as an antiinflammatory in tendon injuries, such as dexamethasone, have proved to be linked to a predisposition for rupture [5]. Recent studies have shown that dexamethasone causes an overall inhibition of cellular proliferation as well as collagen synthesis [6,7]. Novel therapies have suggested that Vitamin C supplementation is not only able to bolster collagen synthesis in the face of injury due to its role in collagen assembly but can also serve as a cytoprotective agent against glucocorticoid induced cellular apoptosis [6,7].



MAIN HYPOTHESIS:

We hypothesize that supplemental vitamin C will bolster collagen synthesis and act as a negative cytoprotective agent against antiinflammatory glucocorticoid in consequences tendon proper (TP) (PERI) peritenon and stem/progenitor cells of the equine superficial digital flexor tendon (SDFT).

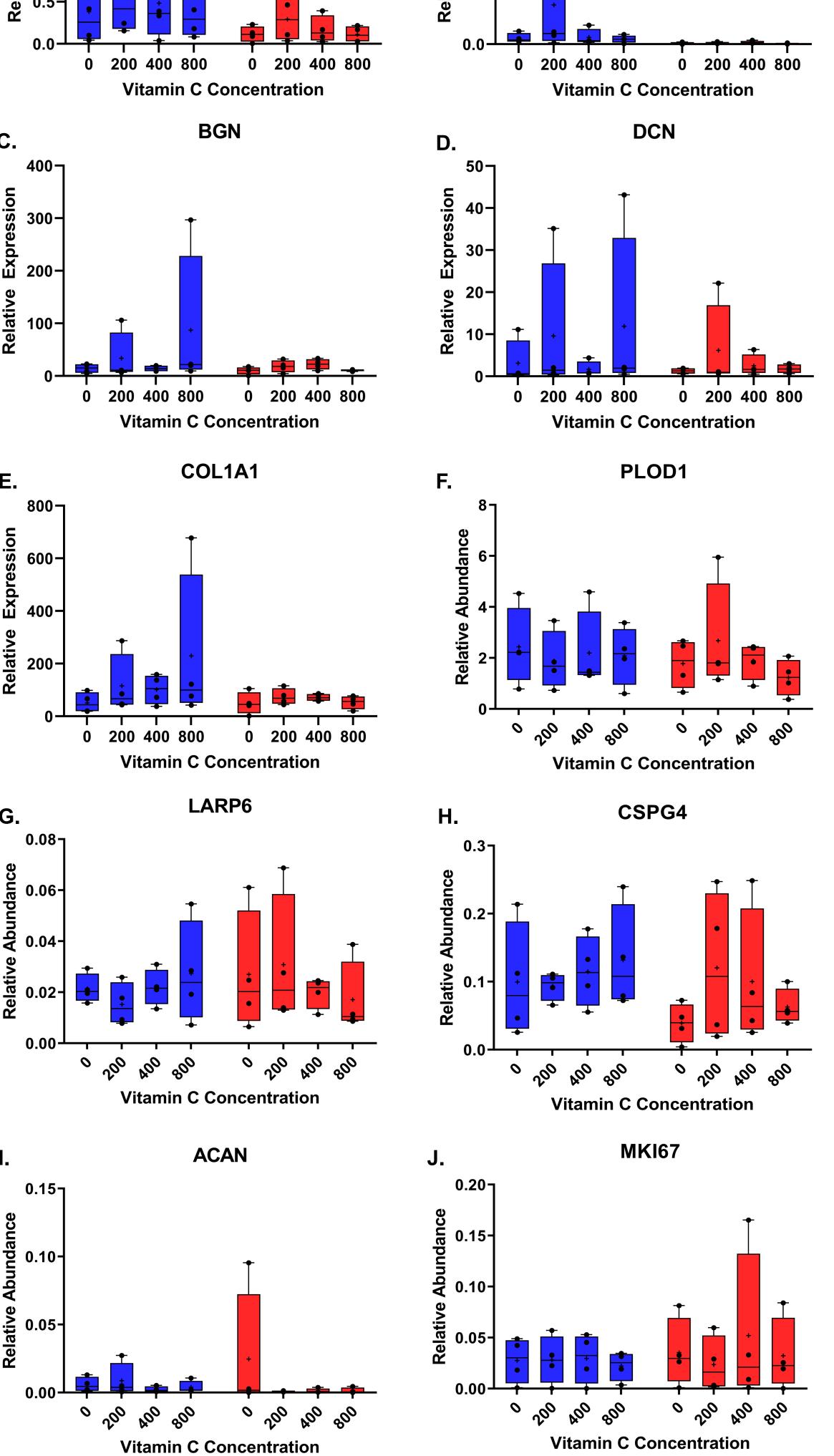
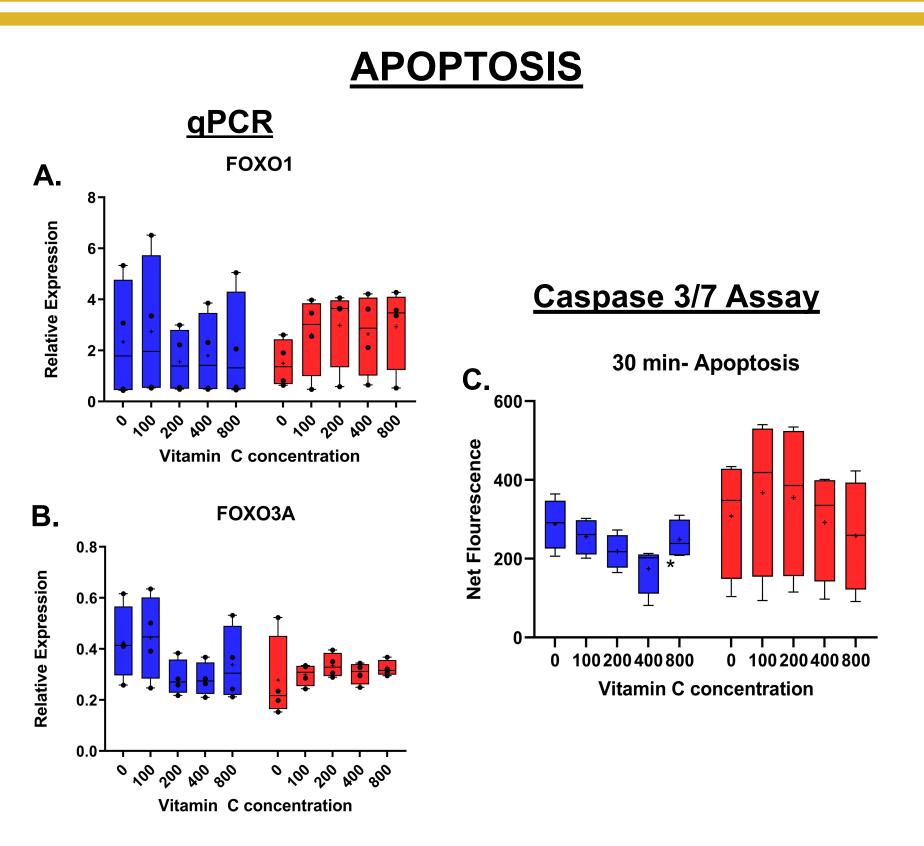


Figure 2: Vitamin C supplementation increases the overall collagen content in tendon constructs with TP and PERI cell populations. (A): Levels of collagen for constructs of TPderived (BLUE) progenitor cells trended upward with increasing amounts of vitamin C. (B): Constructs of PERI-derived (RED) progenitor cells treated with vitamin C showed a significant (p<0.05) increase in collagen content relative to the control in 100 nM, 200 nM, and 400 nM (p<0.01). Statistical significance was found via a paired t-test utilizing GraphPad Prism.



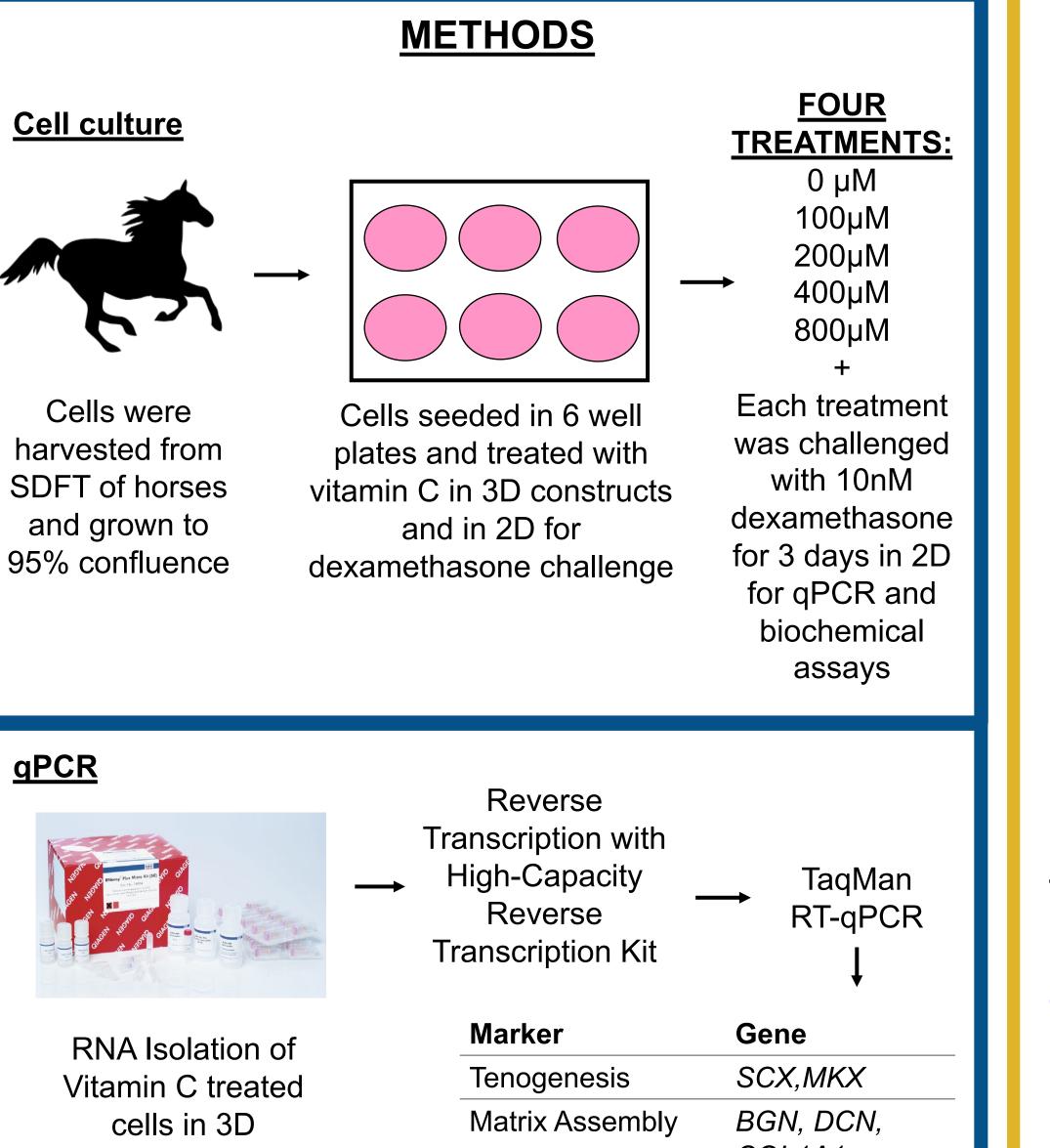


Figure 1: Vitamin C increased the relative expression of tenogenic markers in tendon constructs with TP and PERI-derived cell populations. (A,B): Tenogenic marker SCX trended upwards when supplemented with 200 nM in both cell types. MKX also increased expression when supplemented with 200 nM of vitamin C in TPderived (BLUE) cells but remained unchanged in PERI-derived (RED) cells. (C,D,E,F,G): Matrix assembly markers BGN and DCN showed an increase at concentrations of 400 nM and 800nM in TP cells and no change in PERI cells. COL1A1 increased in expression when supplemented with vitamin C in TP-derived calle whilet PERL derived calls remained unchanged PLOD1 showed no change in

Figure 3: Vitamin C supplementation decreases overall apoptosis in TP and PERI cell populations treated with dexamethasone. (A): Vitamin C supplementation had no effect on the genetic expression of apoptotic marker *FOXO1* in both TPderived (BLUE) and PERI-derived (RED) cells. (B): Apoptotic marker FOXO3A showed a relative decrease in expression mainly in TP-derived cells at 200 nM and 400 nM while expression for PERI-derived cells trended higher, but not with significance. (C): The Apo-ONE Caspase-3/7 assay showed the greatest decrease in apoptotic activity when dexamethasone-challenged TP-derived cells were supplemented with 400 nM of Vitamin C (p<0.05) while PERI-derived cells remained unchanged. Statistics were determined with a paired t-test utilizing GraphPad Prism.

CONCLUSION

vitamin C affected In conclusion, the genetic and PERI 3D TP expression of derived tendon **TP-derived** More specifically, constructs. cells seemed to increase tenogenic genes while PERIderived cells increased both cellular proliferation and

+		COL1A1, PLOD1, LARP6
Vitamin C treated cells challenged with 10 nM treatment dose equivalent of dexamethasone in 2D	Vascular formation	CSPG4
	Chondrification	ACAN
	Cellular proliferation	MKI67
	Apoptosis	FOXO1, FOXO3A
<u>Hydroxyproline Assay</u>	<u>Apoptos</u>	<u>sis Assay</u>
Dried Vitamin C treated 3D tendon constructs in a series of acid and boiling steps Generated L- hydroxyproline standards for standard curve analysis Plated in a clear 96- well plate and read in a spectrophotometer	Caspase-3/7 well of a blac seeded in 2 treatment Dexam Incubated Measured	µl of Apo-ONE Reagent to each ck 96-well plate D after 72 hour with 10 nM of ethasone ↓ for ~30min ↓ fluorescence trophotometer

expression in TP-derived cells but increased in PERI-derived cells supplemented with 200 nM. <i>LARP6</i> showed no change in expression in any cell type or treatment group. (H): Vascular marker <i>CSPG4</i> showed an overall increase in PERI cells compared to the control. (I): Chondrification marker <i>ACAN</i> showed an overall decrease in PERI-derived cells when supplemented with vitamin C while TP-derived cells remained unchanged. (J): Cellular proliferation marker (<i>MK167</i>) showed an increase in expression when supplemented with 400 nM in PERI-derived cells while TP-derived	matrix assembly genes. Vitamin C also caused an overall decrease in apoptotic gene expression as well as apoptotic enzyme activity, mainly for TP cells. From these findings one can conclude that vitamin C not only caused an overall increase in tenogenic effects but also plays a cytoprotective role in dexamethasone challenged cells.
Literature Cited	Acknowledgements
 [1] Zhang J. et al. 2020. Moderate and intensive mechanical loading differentially modulate the phenotype of tendon stem/progenitor cells <i>in vivo</i>. PloS ONE. 15(12):e0242640 [2] Smith R.K.W. and McIlwraith C.W. 2021. "One Health" in tendinopathy research: current concepts. J. Orthop. Res. [3] Hu AJ. and Bramlage LR. 2014. Racing performance of Thoroughbreds with superficial digital flexor tendonitis treated with desmotomy of the accessory ligament of the superficial digital flexor tendon' [4] Tipton TE. et al. Superficial digital flexor tendonitis in cutting horses: 19 cases (2007-2011). Journal of the American Veterinary Medical Association 243:1162-1165 [5] Poulsen R.C., Carr A.J., Hulley P.A. 2011. Protection against Glucocorticoid-Induced Damage in Human Tenocytes by Modulation of ERK, Akt, and Forkhead Signaling. <i>Endocrinology</i> 152(2):503-514 [5] Dean BJF. et al. 2014. The risks and benefits of glucocorticoid treatment for tendinopathy: A systematic review of the effects of local glucocorticoid on tendon. Semin Arthritis Rheum. 2014 Feb;43(4):570-6 [6] Omeroglu S. et al. 2009. High-dose vitamin C supplementation accelerates the Achilles tendon healing in healthy rats. Arch Orthop Trauma Surg. 129(2):281-286 [7]Scutt N., Rolf C.G., Scutt A. 2006. Glucocorticoids Inhibit Tenocyte Proliferation and Tendon Progenitor Cell Recruitment. <i>J Orthop Res</i> 24(2):173-18 	We gratefully acknowledge the financial support from the UC Davis Center of Equine Health, the UC Davis School of Veterinary Medicine, the UC Davis College of Agricultural & Environmental Sciences, the Department of Animal Science at UC Davis, and the Students Training in Advanced Research Program.